Colorimetric Chemosensor for Sulfide Anion Detection Based on Symmetrical Nitrovanillin Azine

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email: purwono.bambang@ugm.ac.id Received: December 15, 2021

Accepted: September 28, 2022 DOI: 10.22146/ijc.71259 **Abstract:** The nitrovanillin azine (**NA**) has been successfully synthesized and examined as a colorimetric chemosensor for sulfide anion detection. The **NA** was synthesized using two steps reaction. Vanillin was reacted with concentrated nitric acid to form 5nitrovanillin (**NV**) then the **NV** was condensed with hydrazine hydrate to produce the **NA**. The **NA** was obtained and fully elucidated by FTIR, ¹H-NMR, ¹³C-NMR, and GC-MS spectrometer. The **NA** activity for anionic chemosensor was then carried out on several anions such as F^- , Cl^- , Br^- , I^- , S^{2-} , CN^- , HCO_3^- , AcO^- , H_2PO4^- , N_3^- , NO_2^- , SCN^- , ClO_3^- , and NO_3^- . The chemosensor tests showed **NA** was only selective for S^{2-} in DMF:HEPES buffer (9:1, v/v, 10 mM, pH = 7.4) giving color change from light yellow to dark green. The LOD value was 1.43×10^{-5} M and the interaction model of **NA**-S²⁻ indicated deprotonation mode between the -OH group with sulfide anion in a ratio of 1:1. The **NA** chemosensor can be applied for qualitatively analysis of sulfide anion using filter paper strips and quantitatively analysis of sulfide anion in tap water.

Keywords: 5-nitrovanillin; hydrazine; azine; colorimetric; chemosensor

INTRODUCTION

Hydrogen sulfide (H₂S) is one of the most toxic, corrosive, and flammable gases with the characteristic odor of rotten eggs. In high concentrations, this gas causes serious problems such as hypertension, diabetes, liver cirrhosis, chronic kidney, Down Syndrome, and Alzheimer's disease [1-3]. According to the National Institute of Occupational Safety and Health, the concentration of H₂S harmful to life or health is up to 150 ppm, and the recommended exposure limit is 10 ppm for a maximum duration of 10 min [4]. Hydrogen sulfide is also involved in pathological and physiological functions, including inhibition of insulin signaling, regulation of inflammation, and relaxation of the smooth muscle of blood vessels [5]. Several methods for detecting sulfides are potentiometry [6], chromatography [7], and electrochemistry [8]. However, the use of those instruments requires a complicated sample preparation, expensive, and difficult to operate. Therefore, it is necessary to develop simple, efficient, and faster methods with a high level of sensitivity and selectivity. The colorimetric chemosensor is the most often used to detect anions through color changes [9]. Visually, the detection process is more profitable because it can provide qualitative and quantitative information in a relatively short time [10], and it is possible to be seen directly with the naked eye. This method is not expensive and easy to operate their instruments [11].

Chemosensor compounds have two main parts: signal site in the form of chromophore groups acting as color carrier such as azine (RHC=N-N=CHR) [12], and binding sites such as -OH playing a role in interacting with anions through hydrogen bonding or deprotonation [13]. Research for anion chemosensor in recent years continues to grow. Acetophenone azine derivative has been synthesized as a colorimetric and fluorescence sensor to detect CN^- , F^- , and AcO^- in DMSO [14]. Azines from vanillin and 2-hydroxy-5-phenylazo-benzaldehyde have been used as a colorimetric sensor of S^{2^-} anion in DMF: HEPES (v/v = 9:1, 10 mM HEPES, pH = 7.4) [15-16]. Another unsymmetrical azine of dicyanoisophoronebased derivatives has been designed as a colorimetric and fluorescence of Cu²⁺ toward S²⁻ in DMSO/HEPES buffer (v/v = 2:1, 20 Mm HEPES, pH = 7.4) [17].

In this study, we synthesized the nitrovanillin azine (NA) from vanillin as a sulfide anion colorimetric chemosensor. Investigation for NA colorimetric chemosensor was carried out by sensing properties measurement to various anion, time interaction, anion interferences, pH effect, reversibility, and LOD. A qualitative test of sulfide anion has also been performed with a filter paper strip, while a quantitative analysis of sulfide anion in tap water shows the application of NA.

EXPERIMENTAL SECTION

Materials

Materials used for the synthesis were vanillin, nitric acid 37%, dichloromethane (DCM), hydrazine hydrate 80%, methanol, ethanol, chloroform, dimethyl sulfoxide (DMSO), dimethylformamide (DMF), HEPES buffer solution (HBS) in deionized water, acetone, acetonitrile, aquadest, sodium fluoride, sodium chloride, sodium bromide, sodium iodide, sodium sulfide, sodium cyanide, sodium hydrogen carbonate, sodium acetate, sodium dihydrogen phosphate, sodium azide, sodium nitrite, potassium thiocyanate, potassium chlorate, and potassium nitrate. All materials used have a Merck p.a quality, while deionized water is from Onemed.

Instrumentation

The characteristics of the synthesized compounds were carried out using a melting point determination apparatus (Electrothermal-9100), FTIR spectrophotometers (Shimadzu Prestige-21), ¹H-NMR (Agilent, 500 MHz and JEOL JNM ECA-500, 500 MHz), ¹³C-NMR (Agilent, 125 MHz), GC-MS and DI-MS (Shimadzu QP-2010S). The color change was measured using a UV-Vis spectrophotometer (Shimadzu UV-1800).

Procedure

Synthesis of 5-nitrovanilin (5-NV)

The 5-nitrovanillin (5-NV) was synthesized based on previous methods [18]. Vanillin (13.79 g, 70 mmol) was dissolved in 55 mL of dichloromethane (0–5 °C) and 12 mL of HNO₃ was added dropwise at 0–5 °C. The reaction mixture was stirred at room temperature for 20 min and then cold distilled water (25 mL) was added and left for 2 h. Subsequently, the solid was filtered and recrystallized with ethanol. The synthesized product was obtained as a pale-yellow powder in yield of 72.1% (m.p. 176–178 °C). FTIR (KBr) (cm⁻¹): 3209 (-OH group), 3078 (Csp²-H), 2887 (Csp³-H), 1681 (C=O), 1543 (-NO₂), 1612 and 1465 (C=C aromatic), 1111 and 1049 (C-O-C). m/z [M⁺] = 197.

Synthesis of nitrovanillin azine (NA)

The synthesis of NA compound is based on a previous method with some modifications [15]. The 5-NV (0.390 g, 1 mmol) was dissolved in 25 mL of ethanol and mixed with (0.5 mmol) hydrazine hydrate 80% in 5 mL of ethanol. The reaction mixture was stirred at room temperature for overnight and the progress of reaction was monitored by TLC. The solid was filtered and washed with ethanol several times and then crystallized with ethanol:DMSO (2:1). The product was obtained as a yellow solid in yield of 76.14% (m.p. 293-394 °C). FTIR (KBr) (cm⁻¹): 3448 (-OH group), 3078 (Csp²-H), 2947 (Csp³-H), 1627 (-C=N-), 1543 (-NO₂), 1472 (C=C aromatic), 1273 (C-O-C). m/z [M⁺] = 390. ¹H-NMR (500 MHz, DMSO- d_6) (δ) (ppm): 3.96 (3H, s, OCH₃), 7.74 (1H, s, ArH), 7.96 (1H, d, ArH), 8.71 (1H, s, -CH=N-), 11.16 (1H, s, ArOH). ¹³C-NMR (δ) (ppm): 56.69 (OCH₃), 112.73 (Caryl), 118.19 (Caryl), 124.10 (Caryl), 137.18 (Caryl-NO2), 145.72 (Caryl-OH), 150.03 (-C=N-), 160.10 (Caryl-OCH₃).

Study of NA selectivity as anion chemosensor

A solution of **NA** $(2 \times 10^{-5} \text{ M})$ was prepared in DMF:HBS (9:1, v/v, 10 mM HEPES, pH = 7.4) and solution of anion (0.2 M) such as F⁻, Cl⁻, Br⁻, I⁻, S²⁻, CN⁻, HCO₃⁻, AcO⁻, H₂PO₄⁻, N₃⁻, NO₂⁻, SCN⁻, ClO₃⁻, and NO₃⁻ in HEPES Buffer Solution (HBS). The 4 mL of **NA** were placed in a cuvette and each (150 µL) anion solution was added. The color change was recorded and determined with a UV-Vis spectrophotometer.

Study of competition of sulfide anions with different anions

A solution of NA $(2 \times 10^{-5} \text{ M})$ was prepared in DMF:HBS (9:1, v/v, 10 mM HEPES, pH = 7.4) and

different anion solutions with a concentration of 0.2 M in HBS. The 4 mL of **NA** were put in a cuvette and different anions (0.2 M, 100 μ L) of F⁻, Cl⁻, Br⁻, I⁻, CN⁻, HCO₃⁻, AcO⁻, H₂PO4⁻, N₃⁻, NO₂⁻, SCN⁻, ClO₃⁻, and NO₃⁻ were added, then into each mixture added S²⁻ (100 μ L). Color changes were recorded and monitored at a wavelength of 613 nm.

Sensitivity study of NA as a sulfide anion sensor

A solution of **NA** $(2 \times 10^{-5} \text{ M})$ was prepared in DMF:HBS (9:1, v/v, 10 mM HEPES, pH = 7.4) and a solution of S²⁻ (0.2 M) in HBS. The 4 mL of **NA** was put in a cuvette and added S²⁻ (0–100 µL). The color change for each addition of S²⁻ was recorded and monitored at a wavelength of 613 nm. Determination of detection limit using the $3\sigma/m$ equation [19].

Study of interaction time of NA-sulfide anions

The 4 mL of **NA** solution $(2 \times 10^{-5} \text{ M})$ was prepared in DMF:HBS (9:1, v/v, 10 mM HEPES, pH = 7.4) and a solution of S^{2–} (0.2 M, 115 µL) was added in HBS. Changes in UV-Vis absorbance were monitored at wavelengths of 613 nm with different time intervals at room temperature.

Reversibility study of NA toward sulfide anions

The 4 mL of **NA** solution $(2 \times 10^{-5} \text{ M})$ was prepared in DMF:HBS (9:1, v/v, 10 mM HEPES, pH = 7.4) and a solution of S²⁻ (0.2 M, 116 µL) was added in HBS followed by the addition of HCl (2 M, 23 µL). Changes in UV-Vis absorbance were monitored at wavelengths of 613 nm before and after adding S²⁻ and HCl several times with different time intervals at room temperature.

pH Dependent study of NA toward sulfide anions

A solution of **NA** $(2 \times 10^{-5} \text{ M})$ was prepared in DMF:HBS (9:1, v/v, 10 mM HEPES, pH = 3, 5, 7.4, 9 and 11) and a solution of S²⁻ (0.2 M) in HBS. The 4 mL of **NA** with various pH was placed in a cuvette and added S²⁻

(150 μ L). Changes in UV-Vis absorbance were monitored at wavelengths of 613 nm before and after the addition of S^{2–}.

Determination of NA-S^{2^-} interaction through Job's Plot

A solution of **NA** in DMF:HBS (9:1, v/v, 10 mM HEPES, pH = 7.4) and solution of S^{2-} in HBS were prepared in a constant concentration of 7×10^{-5} M. Host-Guest solution comparison (1:9–9:1) were prepared by mixing of 0.4–3.6 mL of **NA** solution (Host) with 3.6–0.4 mL of S^{2-} (Guest) with a total solution of 4 mL. Changes in UV-Vis absorbance were monitored at wavelengths of 613 nm and plotted as [G]/[H] vs absorbance.

Filter paper strip study of NA toward sulfide anions

Filter paper strip (Whatman no. 42) with a size of 1×3 cm were immersed in a solution of **NA** 1×10^{-3} M in DMF:HBS (9:1, v/v, 10 mM HEPES, pH = 7.4) for 2 h. The paper strip was dried in an oven, and then two drops of S²⁻ solution (0.2 M) in HBS were added to the paper strip and allowed to react. Color change was recorded visually and under a 366 nm UV lamp.

RESULTS AND DISCUSSION

Chemosensor **NA** was synthesized by condensation reaction between **5-NV** with 80% hydrate hydrazine (Scheme 1) and purified through recrystallization from a solvent mixture of ethanol:DMSO (2:1) to produce a golden yellow solid. All products of synthesis are agreed with spectroscopy data.

Chemosensor **NA** has a binding site in the form of phenol groups that can interact strongly with anions through the hydrogen bond interaction. Here, the -OH group can be deprotonated with strong basic anions such



Scheme 1. Schematic formation of the reaction of NA Chemosensor

as CN⁻, HCO₃⁻, and AcO⁻ [20]. Sulfide anion also has basic properties that allow the same deprotonation mechanism. Hydrogen sulfide (H₂S) dissolves in water and dissociates to form H⁺, HS⁻ and S²⁻. Under physiological conditions, it forms about 20% as H₂S and 80% as HS⁻/S²⁻. In this study, sulfide anion (S²⁻) is a mixture of H₂S + HS⁻ + S²⁻ in solution [21].

Selectivity and Sensitivity of NA toward S²⁻

Chemosensor **NA** in DMF:HBS (9:1, v/v, 10 mM, pH = 7.4) has interacted with different anions such as F⁻, Cl⁻, Br⁻, I⁻, S²⁻, CN⁻, HCO₃⁻, AcO⁻, H₂PO₄⁻, N₃⁻, NO₂⁻, SCN⁻, ClO₃⁻, and NO₃⁻. The color changes were recorded and confirmed using a UV-Vis spectrophotometer. The addition effect of each anion showed that only the S²⁻ anion gave a color change from light yellow to dark green, whereas other anions did not give a significant color change. The absorbance spectra of **NA** showed two peaks at 388 and 459 nm. After addition of S²⁻ showed a bathochromic shift with the appearance of a new absorbance at 613 nm. This shift indicated the interaction of **NA** with S²⁻. The phenol group of **NA** experienced deprotonation and then followed by delocalization of π electrons to produce a color change (Fig. 1) and emerging

wavelength at λ_{max} 613 nm (Fig. 2).

Detection Limit Calculation

The **NA** sensitivity was confirmed using a UV-Vis titration. Changes in UV-Vis absorbance after the addition of S^{2–} (0–5 mM) in DMF:HBS showed an increase in absorbance with increasing S^{2–} concentration at λ_{max} 613 nm (Fig. 3(a)).



Fig 2. UV-Vis absorbance spectra of **NA** $(2 \times 10^{-5} \text{ M})$ with several anions in DMF:HBS (9:1, v/v, 10 mM, pH = 7.4)



Fig 3. (a) Absorbance spectra of **NA** (2×10^{-5} M) after the addition of concentration variations S^{2–} and (b) Linear regression curve of **NA** with S^{2–}

The calculation of the detection limit used equation $3\sigma/m$, where the σ is the standard deviation obtained from an empty signal without S^{2–} and m is the slope value of the calibration curve [19]. The ratiometric calibration curve has an R² of 0.987 (Fig. 3(b)), so the detection limit value (LOD) S^{2–} is 1.43×10^{-5} M (14.3 μ M). This **NA** value is smaller than previous LOD values reported by Kaushik et al. [15] and Rokhmah et al. [16].

Competition Study with Other Anions

Examination of possible interference S^{2-} with other anions such as F⁻, Cl⁻, Br⁻, I⁻, CN⁻, HCO₃⁻, AcO⁻, H₂PO₄⁻, N₃⁻, NO₂⁻, SCN⁻, ClO₃⁻, and NO₃⁻ was studied. The results showed that almost all other anions have no interference effect in detecting S²⁻ anions except for CN⁻ and HCO₃⁻ (Fig. 4). These anions, CN⁻ and HCO₃⁻ are strong bases which interfere sulfide detection process [22]. Sulfide with CN⁻ and HCO₃⁻ competes with each other in interacting with the binding site of the chemosensor. Therefore, there is a slight increase in absorbance. This indicates that S²⁻ is not bound to the **NA** binding site due to the presence of CN⁻ and HCO₃⁻, which causes no color change to occur.

Study of Interaction Time Effect of NA-S²⁻

The study of the interaction time effect of NA-S²⁻ was monitored at λ_{max} 613 nm for 30 min (Fig. 5). The measurement results showed that the absorbance change at the peak of 613 nm occurred within one minute after the addition of S²⁻ and reached a maximum absorbance in 15 min and then decreased over time. The absorbance

decreases occurred because the S^{2–} was in another form after 15 min. This study confirmed that **NA** toward S^{2–} has a reasonably fast response.

Reversibility Study

The study of **NA** reversibility was examined by adding S^{2–} followed by the addition of a dilute acid solution of HCl (2 M) at λ_{max} 613 nm. Absorbance at λ_{max} 613 nm is a form of **NA** deprotonation by S^{2–}, and then after the addition of dilute acid, the absorbance at λ_{max} 613 nm is lost (**NA** is not deprotonated). Based on the absorbance at λ_{max} 613 nm, **NA** chemosensor can be used as a sulfide sensor with repeatability for up to four cycles (Fig. 6). This shows that **NA** has good reversibility to S^{2–} and H⁺ ions.



Fig 5. Interaction time and absorbance relationship curve at λ 613 nm of NA (2 × 10⁻⁵ M) to S²⁻



Fig 4. Bar diagram of the competition of several anions with S^{2–}



Fig 6. Reversibility absorbance spectra of NA with S^{2-} after the addition of HCl (2 M)

pH Effect Study

The pH effect in DMF:HEPES buffer (9:1, v/v, 10 mM HEPES, pH = 3, 5, 7, 4, 9, and 11) was carried out to determine the feasibility of working sensor at different pH. The UV-Vis spectra before the addition of S^{2–} showed two absorptions at 393 and 464 nm for pH of 7.4, 9, and 11. While at pH 3 and 5 produced absorptions at 336 and 337 nm, respectively (Fig. 7(a)). After the addition of S^{2–} at a pH of 7.4 produced the highest maximum absorbance at 613 nm, while at pH 9 and 11, there was no color change and wavelength shifts (Fig. 7(b)).

This shows that **NA** works effectively to detect S^{2-} at a solvent pH 3–8 because the main sulfide species at an acidic pH is H₂S, and pH > 7 is dominant in the form of HS⁻ from the equilibrium of the S^{2–} species from the salt solution which allows to bind to the **NA** binding site [23]. At pH 9–11, H₂S is in the form of HS[–]. The presence of excess HS[–]causes higher basicity so that the sulfide interaction with the **NA** binding site does not occur, similar to the interferent of CN[–] and HCO₃[–].

Job's Plot Study and NMR Titration

This experiment was conducted to determine the stoichiometric ratio of interactions between **NA** (host) and S²⁻ using the Continuous Variant methods (Job's Plot) [24]. The results from Job's Plot curve showed the ratio of 1:1 (Fig. 8). The **NA**-S²⁻ interaction was also studied by ¹H-NMR titration in DMSO- d_6 (Fig. 9). The results showed that the addition of S²⁻ (1 eq.) in D₂O



Fig 8. Job's Plot curve of NA titration toward the addition of $\mathsf{S}^{2\text{-}}$



Fig 7. (a) Absorbance spectra of **NA** (2×10^{-5} M) at different pH and (b) Absorbance spectra of **NA** (2×10^{-5} M) after the addition of S²⁻ to different pH



Fig 9. ¹H-NMR spectra titration of NA with S^{2–}

causes the -OH signal at δ 11.16 ppm to disappear completely, followed by the proton of imine and aromatic ring shifting simultaneously to the up-field area. The addition of S²⁻ (2 eq.) has shifted more up-field than the addition of 1 eq. This shift is caused by -OH deprotonation produced by a resonance phenomenon. The electron density of the aromatic ring increases due to the disconnection of charges in the conjugation system. From the stoichiometric ratio stating one molecule of the binding site (-OH) interacting with one molecule of S²⁻ can be made an interaction model of NA-S^{2–} (Scheme 2).

Application of NA Chemosensor in Tap Water **Samples Analysis**

Determination of % recovery was carried out for quantitative analysis of sulfide in tap water samples. Tap water was added $S^{2-}(1, 3, and 5 \text{ mM})$, and then NA was used as the reagent for quantitative analysis using UV-Vis. Using calibration curves (y = 0.1148x - 0.1216), the % recovery values are 108.5, 98.6 and 91.5%, respectively (Table 1). These results indicate that NA can be used to detect S²⁻ with a concentration range of 1–5 mM. From the obtained analysis data of a range % recovery (80-108%) and % RSD (6%) [25], it can be said that NA can be used for quantitative analysis by UV-Vis spectrophotometer. The results of % recovery and % RSD values are shown in (Table 1).



Scheme 2. The interaction model of NA-S²⁻

I able 1. Determination of sulfides in tap water samples				
Sample	Na ₂ S added	Na ₂ S found	RSD	Recovery
	(mM)	(mM)	(%)	(%)
Tap water	1	1.085 ± 0.02	1.8	108.5
	3	2.958 ± 0.21	3.0	98.6
	5	4.587 ± 0.09	4.6	91.5



Fig 10. Change in the color of paper strip soaked in **NA** $(1 \times 10^{-3} \text{ M})$ after the addition of S^{2–} (0.2 M)

Qualitative Analysis Using Filter Paper Strips

To examine the rapid response of **NA** to S^{2-} , a qualitative analysis was performed using filter paper. Whatman filter paper No. 42 immersed in NA solution $(1 \times 10^{-3} \text{ M})$, was added two drops of S^{2-} and dried (Fig. 10). The color of paper strip changes from light pink to yellow. Furthermore, the paper strip was monitored under a 366 nm UV lamp and indicated no luminescence both before and after the addition of S^{2-} . The results indicated that **NA** could be used to detect sulfides in colorimetric analysis.

CONCLUSION

We have successfully synthesized a nitrovanillin azine (NA) through the condensation reactions of 5nitrovanillin (5-NV) with hydrazine hydrate. Properties of NA chemosensor gave high selectivity toward S²⁻ in DMF:HBS (9:1, v/v, 10 mM, pH = 7.4) with color change from light yellow to dark green. The colorimetric LOD of NA was 14.3 μ M, and the interaction model between host-guests (NA-S²⁻) is in a 1:1 ratio. A filter paper strip for qualitative and quantitative analysis of S²⁻ in tap water samples indicated that NA has potential as a sulfide anion colorimetric chemosensor.

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