The Effect Of MSC-Conditioned Medium (MSC-CM) Low Dosage on Urea Levels in Acute Renal Failure Rats

Rizqi Windhu Sri Intania^{1*}, Azizah Retno Kustiyah², Nur Anna Chalimah Sa'dyah³

* Correspondence: rizqiwindhu@std.unissula.ac.id

¹Student of the Faculty of Medicine, Universitas Islam Sultan Agung, Semarang, Indonesia
² Department of Pediatrics, Faculty of Medicine, Universitas Islam Sultan Agung, Semarang, Indonesia
² Department of Internal Medicine, Faculty of Medicine, Universitas Islam Sultan Agung, Semarang, Indonesia

Received 08 July 2022 Accepted 29 August 2022 Available online on 30 September 2022

© 2022 The Authors. Published by Stem Cell and Cancer Research, Semarang, Indonesia. This is an open-access article under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike License (CC BY-NC-SA 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Background: Acute renal failure (ARF) is associated with a high incidence of morbidity and mortality, as well as a high risk of developing chronic renal failure that is associated with elevation of urea level. The kidneys have an extraordinary ability to regenerate after injury and fully recover, and clinical options are limited to fluid management and dialysis procedures. The development of new strategies to increase the ability of kidney regeneration due to ARF, and to maintain kidney function both in the short term and in the long term is needed.

Objective: This study aims to determine the effect of low-dose MSC-CM on urea levels in ARF.

Method: In this study, 12 male gentamicin-induced Wistar rats were divided into 2 groups control (intravenous PBS administration) and treatment (intravenous MSC-CM low-dose at 0,2 cc). Parameters including urea-urine level were measured spectrophotometer and then analyzed by unpaired T-test.

Result: The results of this study showed that the mean urea levels between the control group (19.46 \pm 0.56 mg / dL) and the treatment group (13.96 \pm 0.73 mg / dL) were significantly different (p <0.05).

Conclusion: The conclusion of this study indicated that there was an effect of low-dose of MSC-CM on urea levels in acute renal failure.

Keyword : Acute Renal Failure, MSC-CM, Urea Level

INTRODUCTION

Acute Renal Failure (ARF) still has a high incidence of morbidity and mortality and is at high risk of developing into chronic failure^{1–3}. One of the assessments of kidney function is an increase in serum urea in the body^{4,5}. High levels of urea can be used as an indicator of ARF². There are two treatment options for kidney failure include dialysis (hemodialysis or peritoneal dialysis) and kidney transplantation that have side effect due to the body's immune system for a lot of different reasons⁶. It is important to realize that transplant patients have no control over most of these causes of transplant failure⁷. While the low blood pressure (hypotension) is one of the most common side effects of hemodialysis⁸.

The development of new strategies in order to increase the ability to regenerate after ARF, as well as to maintain kidney function both in the short and long term is urgently needed^{1,9}. Stem cell-based therapy is a promising option where this therapy requires cells with maximum ability to support cardiac regeneration^{10,11}. The protective effects on organs by mesenchymal stem cells (MSC) and

and mesenchymal stem cell conditioned medium (MSC-CM) has been enhanced in recent decades, suggesting that either MSC or MSC-CM are capable of enhancing tubular regeneration through their paracrine effects^{12,13}. However, until now, there are not many publications related to the effect of MSC-CM on urea in acute renal failure.

Acute Renal failure (ARF) is a major public health problem, it is associated with a high mortality rate, morbidity, and long-term risk of becoming a chronic disease^{1,14}. In some developed countries, the prevalence of ARF is increasing³. About 15% of patients are hospitalized due to ARF and most of them are in critical condition^{2,3}. In critically ill patients, the rate of ARF varies according to the population studied and the definition of ARF used is around 8–89% in pediatric patients and 7–25% in adults^{13,14}. The Assessment of Worldwide Acute Kidney Injury, Renal Angina, and Epidemiology (AWARE) found a relationship between the incidence of ARF with morbidity and mortality in patients aged 3 months to 25 years based in the intensive care unit¹⁴. Cases of ALF if not handled properly, may give unfavorable results, besides that it can increase hospital stay, can develop into chronic home disease, and can increase mortality^{5,15}. Based on several research references, in rat models of acute and chronic renal failure, the MSC-CM dose used was 0,2 cc as the lowest dose and 0,4 cc as the highest dose¹⁵.

The paracrine effects of MSCs can also be produced by MSC-CM. This condition allows MSC-CM to be used for therapy as a substitute for MSC itself. MSC-CM contains various products of all bioactive molecules such as growth factors secreted by MSCs in culture media¹⁶. MSC-CM contains many growth factors such as IGF-1, TGF- β 1, HGF, and VEGF¹⁷. MSC-CM therapy known was able to increase the regeneration of injured kidney tissue by suppressing cell infiltration reducing interstitial fibrosis and reducing changes in the structure of the glomerulus¹⁸. In vitro research conducted by Moghadasali et al. (2012) used the culture of proximal tubular epithelial cells, which are cells in the kidney induced by gentamicin, which were obtained as a result of cell repair after administration of MSC-CM from the bone marrow of MSCs¹⁹. Several studies have also shown that MSC-CM can improve podocyte apoptosis in STZ-induced type 1 diabetes models and provide glomerular repair in tissues in chronic disease models^{20,21}. This study aimed to evaluate the efficacy of treatment with conditioned medium-mesenchymal stem cells (MSC-CM) in an acute renal failure model in rats of low doses of urea levels.

MATERIAL AND METHODS

Ethical Clearance

This research was conducted at the Stem Cell and Cancer Research (SCCR) Laboratory, Faculty of Medicine, Sultan Agung Islamic University, Semarang. The population in this study were male Sprague Dawley rats kept in the enclosure of the Anatomical Pathology Laboratory, Faculty of Medicine, Universitas Islam Sultan Agung with number 254/VII/2021/Komisi Bioetik.

Acute Renal Failure Animal Model

The sample size in this study was 12 rats, with 6 rats in each group. In this study, 2 groups were used where rats were induced by gentamicin at a dose of 60mg/kg BW intraperitoneally for 10 days. Furthermore, 3 rats were terminated and their kidney histopathology was examined to assess kidney damage. Each group became 5 rats. Furthermore, the control group was treated with PBS intravenously, while the treatment group received an injection of MSC-CM at a low dose of 0.2 cc intravenously.

Urine Collection and Urea Spectrophotometry Analysis

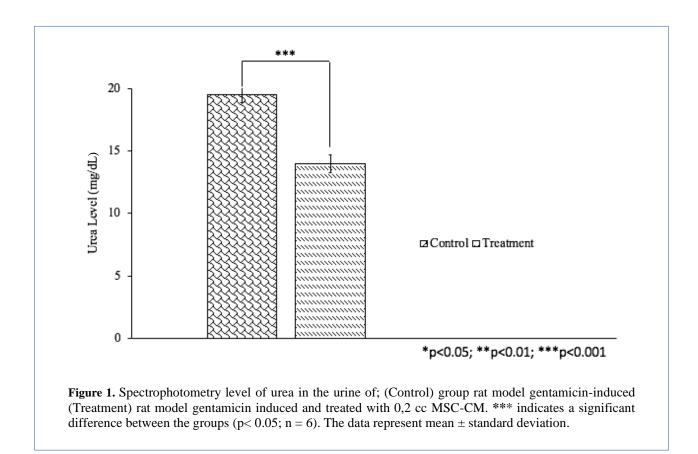
All treatments were carried out on the 11th day. Urea levels in urine were measured in each group after the 19th day. The concentration of the sample and standard against the blank was read using spectrophotometry. In units of mg/dL.

Statistical Analysis

Furthermore, the data were obtained, processed, edited, tabulated, and cleaned, then carried out descriptive data using the mean, median, and mode. Then test the normality of the data using the Shapiro-Wilk test. The data was obtained normally distributed so that the independent sample t-test was carried out differently. Processing of data analysis was carried out using SPSS 20.0 for Windows.

RESULTS

The results of this study showed that the mean urea levels between the control group (19.46 \pm 0.56 mg/dL) and the treatment group (13.96 \pm 0.73 mg/dL) were significantly different (Figure 1). The results of descriptive statistical tests in this study decreased the average level of urea in the treatment group compared to the control group. Based on the results of the independent sample T-test, (p-value = 0.000) which indicates a significant difference in the average urea level in the treatment group (Treatment) compared to the control group (Control). This shows that there is an effect of giving high doses of MSC-CM on urea levels in acute kidney failure compared to controls.



DISCUSSION

In this study, it was found that the administration of low-dose MSC-CM (0.2 cc) was able to significantly reduce urea levels compared to the control group. Several other studies are similar to the results of this study²²⁻²⁴. In this study, compared with the administration of MSC-CM in mice induced by acute kidney failure with cisplatin, the BUN and serum creatinine levels significantly decreased in the negative control group. Research conducted by Birtwistle et al. (2021) explained that the administration of MSC-CM both induced under hypoxic conditions and in normal culture was able to reduce kidney damage parameters, namely serum creatinine in the AKI model induced by cisplatin compared to the control group¹⁴. Research conducted by Tsuji et al. (2018) proved that presenting MSC-CM can improve kidney function in rats with acute renal failure models by reducing the rate of tissue damage and tubular cell apoptosis¹³. Different results were found in the research conducted by Zhao et al. (2020) explained that a significant decrease in BUN levels occurred in the MSC treatment group and not the MSC-CM group in the AKI rat model induced by ischemia-reperfusion injury²⁵.

Gentamicin is an aminoglycoside antibiotic that is widely prescribed to effectively treat patients with infections, but the associated side effects of oxidative stress and injury limit its long-term use². The onset of renal failure is usually slower and the daily increase in serum creatinine tends to be lower than for other causes of acute renal failure³. Serum creatinine and blood urea nitrogen typically increase 7-10 days after initiation of aminoglycoside therapy⁵. Gentamicin as an aminoglycoside drug can cause damage to renal tubular cells such as disruption of the lysosomal membrane structure, mitochondria, and plasma²⁶. Gentamicin-induced nephrotoxicity is characterized by direct tubular necrosis, which is localized mainly in the proximal tubular region²⁷.

Mesenchymal stem cell conditioned medium (MSC-CM) is a product development of stem cells that utilize the secretome of MSC to take advantage of the role of conventional MSC therapy in healing various tissue damage²⁸. Several contents of MSC-CM based on proteomic studies gave the results that IGF-1 levels were 1515.6 ± 211.8 pg/mL; VEGF levels 465.8 ± 108.8 pg/mL; TGF- β 1 levels were 339.8 ± 14.4 pg/mL and HGF levels were 20.3 ± 7.9 pg/mL²⁹. In the process of repairing kidney injury, VEGF becomes an additional factor in renoprotection, while IGF-1 and HGF play a role in the acute regeneration of the post-injured kidney³⁰. Exposure to gentamicin causes necrosis of the proximal tubule, thus giving MSC- In vitro CM in human kidney cell culture has been shown to increase tissue repair by increasing cell viability and accelerating cell migration after gentamicin-induced cell toxicity²³.

MSC-CM significantly reduced tubular cell apoptosis, improved kidney function, and increased survival in mice suffering from acute kidney injury^{19,29,30}. MSC-CM efficiently produces nephrotoxicity by reducing the influx and capacity of dendritic cells and T lymphocyte inflammatory cytokines^{31,32}. In addition, the role of nitric oxide is responsible for the MSC-CM-mediated reno-protective effect^{33,34}. MSC-CM injection significantly increased cisplatin-induced acute kidney injury and inflammation as indicated by decreased levels of serum creatinine, IL-1, and IL-6 and the presence of activated neutrophils in the injured kidney^{21,35,36}. This study indicates that low-dose MSC-CM affects improving kidney function compared to the control group which is in line with previous researches ^{32,37,38}. At the pre-clinical stage for experimental animals, presenting MSC-CM was able to provide improvement in kidney function parameters damaged by gentamicin induction^{22,39,40}. The constraint of this research is the MSC isolation technique to obtain MSC-CM which is so complex^{33,41,42}. This study has limitations where the researcher did not analyze the involvement of various molecules found by MSC-CM in this study, so the regeneration mechanism in this study still refers to the theory or previous studies. There was no examination of the histology of the tissue, such as the tubular injury score so tissue repair was only seen from the functional function of the kidney through a decrease in urea levels.

CONCLUSION

Administration of MSC-CM can reduce urea levels in acute renal failure rat models. Further research can be explored further by assessing the tubular injury score after MSC-CM administration in rat models of acute kidney failure with various doses

FUNDING

None

ACKNOWLEDGEMENTS

This research was supported by the Stem Cell and Cancer Research Laboratory.

AUTHORS' CONTRIBUTIONS

R.W.S.I. conducted the majority of the experiments, analyzed data, and prepared the manuscript. A.R.K. conducted data analysis and provide suggestions for the project. N.A.C.S. supervised the project, conceived the experiments, and wrote most of the manuscript.

COMPETING INTERESTS

The authors declare that there was no conflict of interest.

REFERENCES

- 1. Rayego-Mateos S, Marquez-Expósito L, Rodrigues-Diez R, et al. Molecular Mechanisms of Kidney Injury and Repair. *Int J Mol Sci.* 2022;23(3):1-20. doi:10.3390/ijms23031542
- 2. Putra A, Pertiwi D, Milla MN, et al. Hypoxia-preconditioned MSCs have superior effect in ameliorating renal function on acute renal failure animal model. *Open Access Maced J Med Sci.* 2019;7(3):305-310. doi:10.3889/oamjms.2019.049
- 3. Putra A. Hypoxia-preconditioned MSCs Have Superior Effect in Ameliorating Renal Function on Acute Renal Failure Animal Model. Published online 2019:1-6.
- 4. Kumar S, Verma R, Tyagi N, Gangenahalli G, Verma YK. Therapeutics effect of mesenchymal stromal cells in reactive oxygen species-induced damages. *Hum Cell 2021 351*. 2021;35(1):37-50. doi:10.1007/S13577-021-00646-5
- 5. Xianyuan L, Wei Z, Yaqian D, et al. Anti-renal fibrosis effect of asperulosidic acid via TGF-β1/smad2/smad3 and NF-κB signaling pathways in a rat model of unilateral ureteral obstruction. *Phytomedicine*. 2019;53:274-285. doi:10.1016/j.phymed.2018.09.009
- 6. Aleksandrova SA, Nashchekina YA, Nadezhdin S V, et al. Russian Text © The Author(s), 2020. *Cell tissue biol*. 2020;14(6):238-249. doi:10.1134/S1990519X20060024
- Goto K, Takemura G, Takahashi T, et al. Intravenous Administration of Endothelial Colony-Forming Cells Overexpressing Integrin β 1 Augments Angiogenesis in Ischemic Legs. Stem Cells Transl Med. 2016;5(2):218-226. doi:10.5966/sctm.2015-0096
- 8. Sirivarasai J, Kaojarern S, Chanprasertyothin S, et al. Environmental lead exposure, catalase gene, and markers of antioxidant and oxidative stress relation to hypertension: An analysis based on the EGAT study. *Biomed Res Int*. 2015;2015. doi:10.1155/2015/856319
- 9. Liu Y, Su YY, Yang Q, Zhou T. Stem cells in the treatment of renal fibrosis: a review of preclinical and clinical studies of renal fibrosis pathogenesis. *Stem Cell Res Ther*. 2021;12(1):1-18. doi:10.1186/s13287-021-02391-w
- 10. Huang YZ, Gou M, Da LC, Zhang WQ, Xie HQ. Mesenchymal Stem Cells for Chronic Wound Healing: Current Status of Preclinical and Clinical Studies. *Tissue Eng Part B Rev.* 2020;26(6):555-570. doi:10.1089/TEN.TEB.2019.0351/ASSET/IMAGES/LARGE/TEN.TEB.2019.0351_FIGURE1.JPEG
- 11. Liu W, Yu M, Xie D, et al. Melatonin-stimulated MSC-derived exosomes improve diabetic wound healing through regulating macrophage M1 and M2 polarization by targeting the PTEN / AKT pathway. Published online 2020:1-15.

- 12. Putra A, Rosdiana I, Darlan DM, et al. Intravenous Administration is the Best Route of Mesenchymal Stem Cells Migration in Improving Liver Function Enzyme of Acute Liver Failure. *Folia Med (Plovdiv)*. 2020;62(1):52-58. doi:10.3897/folmed.62.e47712
- 13. Tsuji K, Kitamura S, Wada J. Secretomes from mesenchymal stem cells against acute kidney injury: Possible heterogeneity. *Stem Cells Int.* 2018;2018. doi:10.1155/2018/8693137
- 14. Birtwistle L, Chen XM, Pollock C. Mesenchymal stem cell-derived extracellular vesicles to the rescue of renal injury. *Int J Mol Sci.* 2021;22(12). doi:10.3390/ijms22126596
- 15. Ruiz-Ortega M, Rayego-Mateos S, Lamas S, Ortiz A, Rodrigues-Diez RR. Targeting the progression of chronic kidney disease. *Nat Rev Nephrol*. 2020;16(5):269-288. doi:10.1038/s41581-019-0248-y
- 16. Ho CH, Lan CW, Liao CY, Hung SC, Li HY, Sung YJ. Mesenchymal stem cells and their conditioned medium can enhance the repair of uterine defects in a rat model. *J Chinese Med Assoc.* 2018;81(3):268-276. doi:10.1016/j.jcma.2017.03.013
- 17. Hsiao STF, Asgari A, Lokmic Z, et al. Comparative analysis of paracrine factor expression in human adult mesenchymal stem cells derived from bone marrow, adipose, and dermal tissue. *Stem Cells Dev.* 2012;21(12):2189-2203. doi:10.1089/scd.2011.0674
- Li Z, Wei H, Deng L, Cong X, Chen X. Expression and secretion of interleukin-1β, tumour necrosis factor-α and interleukin-10 by hypoxia- and serum-deprivation-stimulated mesenchymal stem cells. *FEBS J*. 2010;277(18):3688-3698. doi:10.1111/j.1742-4658.2010.07770.x
- 19. Gunawardena TNA, Rahman MT, Abdullah BJJ, Kasim NHA. Conditioned media derived from mesenchymal stem cell cultures: The next generation for regenerative medicine. *J Tissue Eng Regen Med.* 2019;13(4):569-586. doi:10.1002/TERM.2806
- 20. Hashemi SM, Zuhair ·, Hassan M, et al. Investigating the route of administration and efficacy of adipose tissuederived mesenchymal stem cells and conditioned medium in type 1 diabetic mice. *Inflammopharmacology*. 123AD;28:585-601. doi:10.1007/s10787-019-00661-x
- 21. Al-Azzawi B, McGuigan DH, Koivula FNM, et al. The Secretome of Mesenchymal Stem Cells Prevents Islet Beta Cell Apoptosis via an IL-10-Dependent Mechanism. *Open Stem Cell J.* 2020;6(1):1-12. doi:10.2174/1876893802006010001
- 22. Tsuji K, Kitamura S, Wada J. Immunomodulatory and regenerative effects of mesenchymal stem cell-derived extracellular vesicles in renal diseases. *Int J Mol Sci.* 2020;21(3). doi:10.3390/ijms21030756
- Lv S, Liu G, Sun A, et al. Mesenchymal stem cells ameliorate diabetic glomerular fibrosis in vivo and in vitro by inhibiting TGF-β signalling via secretion of bone morphogenetic protein 7. *Diabetes Vasc Dis Res.* 2014;11(4):251-261. doi:10.1177/1479164114531300
- 24. Zhou T, Li HY, Liao C, Lin W, Lin S. Clinical Efficacy and Safety of Mesenchymal Stem Cells for Systemic Lupus Erythematosus. *Stem Cells Int*. 2020;2020. doi:10.1155/2020/6518508
- 25. Zhao X, Zhao Y, Sun X, Xing Y, Wang X, Yang Q. Immunomodulation of MSCs and MSC-Derived Extracellular Vesicles in Osteoarthritis. *Front Bioeng Biotechnol*. 2020;8(October):1-14. doi:10.3389/fbioe.2020.575057
- 26. Fard RMN, Barton MD, Heuzenroeder MW. Bacteriophage-mediated transduction of antibiotic resistance in enterococci. *Lett Appl Microbiol*. 2011;52(6):559-564. doi:10.1111/J.1472-765X.2011.03043.X
- 27. Ghanayem NM, El-shafie MK, Badr EAF, et al. Study of the heat shock protein 70-1 gene polymorphism and the risk of nephropathy in type II diabetic patients. *Menoufia Med J.* 2014;1:582-588. doi:10.4103/1110-2098.145519
- Balasubramanian S, Thej C, Walvekar A, et al. Evaluation of the Secretome Profile and Functional Characteristics of Human Bone Marrow Mesenchymal Stromal Cells-Derived Conditioned Medium Suggest Potential for Skin Rejuvenation. J Cosmet Dermatological Sci Appl. 2017;07(01):99-117. doi:10.4236/jcdsa.2017.71010
- 29. Bhaskar V, Konala R, Bhonde R, Pal R. Secretome studies of mesenchymal stromal cells (MSCs) isolated from three tissue sources reveal subtle differences in potency. doi:10.1007/s11626-020-00501-1/Published
- 30. Li T, Liu Y, Yu L, et al. Human Umbilical Cord Mesenchymal Stem Cells Protect Against SCA3 by Modulating the Level of 70 kD Heat Shock Protein. *Cell Mol Neurobiol*. 2018;38:641-655. doi:10.1007/s10571-017-0513-1
- 31. Saheli M, Bayat M, Ganji R, et al. Human mesenchymal stem cells-conditioned medium improves diabetic wound healing mainly through modulating fibroblast behaviors. *Arch Dermatol Res.* 2020;312(5):325-336. doi:10.1007/s00403-019-02016-6

- 32. Lai P, Weng J, Guo L, Chen X, Du X. Novel insights into MSC-EVs therapy for immune diseases. *Biomark Res.* 2019;7(1):1-10. doi:10.1186/s40364-019-0156-0
- 33. Kupcova Skalnikova H. Proteomic techniques for characterisation of mesenchymal stem cell secretome. *Biochimie*. 2013;95(12):2196-2211. doi:10.1016/J.BIOCHI.2013.07.015
- 34. Putra A, Alif I, Hamra N, et al. MSC-released TGF-β regulate α-SMA expression of myofibroblast during wound healing. *J Stem Cells Regen Med*. 2020;16(2):73. doi:10.46582/JSRM.1602011
- 35. Nugraha A, Putra A. Tumor necrosis factor-α-activated mesenchymal stem cells accelerate wound healing through vascular endothelial growth factor regulation in rats. Universa Med. 2018;37(2):135. doi:10.18051/univmed.2018.v37.135-142
- 36. Kucharzewski M, Rojczyk E, Wilemska-Kucharzewska K, Wilk R, Hudecki J, Los MJ. Novel trends in application of stem cells in skin wound healing. *Eur J Pharmacol*. 2019;843:307-315. doi:10.1016/j.ejphar.2018.12.012
- 37. Putra A, Ridwan FB, Putridewi AI, et al. The Role of TNF-α Induced MSCs on Suppressive Inflammation.pdf. 2018;6(10):1779-1783.
- 38. Ohashi CM, Caldeira FAM, Feitosa DJS, et al. Stem cells from adipose tissue improve the time of wound healing in rats. *Acta Cir Bras*. 2016;31(12):821-825. doi:10.1590/S0102-865020160120000007
- 39. Isakson M, De Blacam C, Whelan D, McArdle A, Clover AJP. Mesenchymal Stem Cells and Cutaneous Wound Healing: Current Evidence and Future Potential. *Stem Cells Int*. 2015;2015. doi:10.1155/2015/831095
- 40. Lo ZJ, Lim X, Eng D, et al. Clinical and economic burden of wound care in the tropics: a 5-year institutional population health review. *Int Wound J.* 2020;17(3):790-803. doi:10.1111/iwj.13333
- 41. Kementerian Kesehatan RI. Riset Kesehatan Dasar Tahun 2018.; 2018.
- 42. Hamra, N. F., Putra, A., Tjipta, A., Amalina, N. D., & Nasihun, T. (2021). Hypoxia mesenchymal stem cells accelerate wound closure improvement by controlling α-smooth muscle actin expression in the full-thickness animal model. *Open Access Macedonian Journal of Medical Sciences*, 9(A), 35-41.